



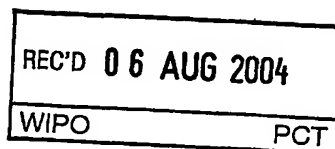
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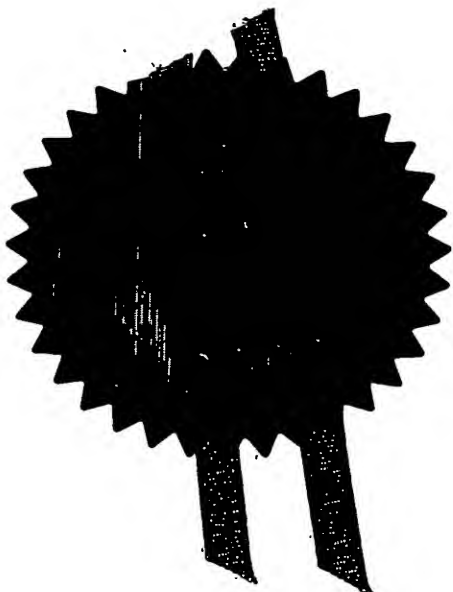
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G139.009.00

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20 MAY 2003

0311565.6

3. Full name, address and postcode of the or of each applicant *(under the off surname)*Cellpep S.A.  
16 rue de la Banque  
75002 Paris  
FrancePatents ADP number *(if you know it)*

If the applicant is a corporate body, give the country/state of its incorporation

French

7560113002

4. Title of the invention

Modified antiviral peptides with increased activity and cell membrane affinity

5. Name of your agent *(if you have one)*"Address for service" in the United Kingdom to which all correspondence should be sent *(including the postcode)*SERJEANTS  
25 The Crescent  
King Street  
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1461001

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**TITLE**

Modified antiviral peptides with increased activity and cell membrane affinity

**DESCRIPTION**

The invention relates to compounds with increased antiviral activity, in particular increased anti-HIV activity, due to the covalent graft on the original antiviral molecule of a structure capable of cell membrane interaction and/or crossing.

**Background**

Multiple branch peptide constructions (MBPCs) comprise a core matrix to which small peptides are bonded. The core matrix is a dendritic polymer which is branched in nature, preferably with each of the branches thereof being identical. Although other core molecules are possible, the preferred core molecule is lysine. The core matrix can be built up from a central lysine residue, sometimes called the root of the MBPC. Two lysine residues are bonded to the central lysine residue, each through its carboxyl group to a different one of the amino groups of the central lysine residue. This provides a molecule with four amino groups, which may be the core matrix for an MBPC having four peptides. Alternatively by bonding a further four lysine residues, each through its carboxyl group to a different one of the said four amino groups, one can provide a molecule with eight branches. This molecule can serve as the core matrix for an MBPC having eight peptides or can alternatively receive eight lysine residues in the manner described above to form a core matrix for an MBPC having sixteen peptides. The C-ends of peptides are covalently bonded to each of the branches of the core matrix to form the MBPC. The peptides may be the same, which is preferred, or may be different from one another. The resulting molecule has a cluster of peptides at the surface and an interior core matrix which is not presented and is therefore not antigenic.

Spacers may, if desired, be included between the peptides and the core matrix. The carboxyl group of the first lysine residue may be left free, amidated, or coupled to a blocking compound such as  $\beta$ -alanine ( $\beta$ -aminopropionic acid). Peptides can include D or L-amino acid residues. D amino acids last longer in vivo because they are harder for peptidase to cut, but the L amino acids have better activity. Moreover, peptide analogues, synthetic constructs using the carbon skeleton of peptides but omitting the -CONH- peptide bonds, can be employed in place of peptides. Thus, it should be understood that references to peptides

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herein may also be taken to include peptide analogues. It is believed that peptide analogues will be more resistant to peptidase and last longer in vivo. If the peptide is too long, the MBPC will become antigenic. It is therefore desirable that each peptide should have not more than ten, and preferably not more than nine, amino acid residues.

MBPCs for use in the treatment of HIV infections were first described by J-M. Sabatier et al in WO 95/07929. The MBPCs described therein have peptides which contain the sequence GPGR (from the V3 loop of the surface envelope glycoprotein gp120 of HIV) preceded by from 0 to 4 amino acid residues and succeeded by from 2 to 4 amino acid residues. The amino acid sequences IGPGR and IXXGPGR (where X is an amino acid residue) are excluded. The most preferred of these MBPCs has a lysine residue core with eight peptides GPGRA<sub>F</sub> bonded thereto. It may be represented as (GPGRA<sub>F</sub>)<sub>8</sub>-K<sub>4</sub>-K<sub>2</sub>-K-βA-OH, the OH terminal indicating the carboxyl group of the β-alanine. That carboxyl group may alternatively be modified to form a carboxamide terminal. This compound is referred to herein as SPC3.

In WO 98/29443, J-M Sabatier et al described further MBPCs which may be effective in the treatment of HIV infection. These use peptides derived from the HIV envelope transmembrane glycoprotein gp41. The peptides contain the sequence RQGY preceded by from 0 to 4 amino acid residues and succeeded by from 2 to 4 amino acid residues. The most preferred of these MBPCs has a lysine residue core with eight peptides RQGYSP<sub>L</sub> bonded thereto. It may be represented as (RQGYSP<sub>L</sub>)<sub>8</sub>-K<sub>4</sub>-K<sub>2</sub>-K-βA-OH, the OH terminal indicating the carboxyl group of the β-alanine. That carboxyl group may alternatively be modified to form a carboxamide terminal. This compound is referred to herein as RL, although it has in the past also been referred to as SPC RL and as RL41.

Subsequently to WO 98/29443, it was established that the MBPC (RQGYSP<sub>L</sub>)<sub>2</sub>-K-βA (hereinafter RL dimer) is effective but that the MBPC (RQGYSP)<sub>2</sub>-K-βA is less so. This was thought to confirm the lower limit of 6 amino acids in the peptide branches of the MBPCs. However, K Mabrouk et al showed in PCT/E<sup>9</sup>03/04353 that some shorter peptides could be used, in particular (RQGYSP)<sub>2</sub>-K-βA-OH (hereinafter RS, but in the past also referred to in the past as Short RL) and (RQGY)<sub>8</sub>-K<sub>4</sub>-K<sub>2</sub>-K-βA-OH.

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SPC3 and RJ. both have 8 branches and are described as octomers. RS has two branches, and is described as a dimer. None of the monomers, that is the linear peptides GPGRAF, RQGYSP and RQGYS, has ever shown any activity.

Anti HIV agents such as SPC3 and RL have been shown to block the fusion step of retroviral infection through direct interaction with cell membrane receptors; other anti fusion agents such as enfuvirtide and T-1249 (Trimeris Inc) interact directly with the viral envelope glycoproteins. The activity of the latter depends on the structure of such glycoproteins, and therefore on the viral strain. Ultimately, molecules that interfere directly with viral glycoproteins will lead to the selection of resistant strains. On the contrary, molecules which are able to block cell membrane receptors should not lead to viral selection, as all strains will be similarly inhibited.

Cell receptor blocking HIV inhibitors may interact with the surface of such receptors (for instance CXCR4 or CCR5) but also with intra membrane components of said receptors, or even with sub-membrane sites or events.

As an example, SPC3, which is an extremely water-soluble peptide, has an anti HIV activity *in vitro* on C8166 cultured cells as well as on peripheral blood lymphocytes (PBL) and on macrophages. B de Rouge in WO 99/34777 showed that this activity is increased 5 to 50 times when SPC3 is associated with certain types of liposomes, probably because of better interaction with cell membranes. However, SPC3 is a polymerized peptide of 56 amino-acid residues. Its association with liposomes is difficult and the yield is not perfect, leading to cost increases as well as technical risks. Other means of improving the efficacy of molecules like SPC3 have therefore been sought.

#### **The invention**

The invention provides a compound comprising a water soluble antiviral peptide and, bonded to the C-end thereof, a terminator which is either (a) an  $\omega$ -amino-fatty acid having from 4 to 10 carbon atoms and from 0 to 2 carbon-carbon double bonds or (b) a peptidic cell membrane penetrating agent.

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The antiviral peptide may be an MBPC with a lysine core matrix. In such a case the terminator is bonded to the root lysine residue. The MBPCs described above may be used, that is to say SPC3 which has 8 branches of GPGRF, RL which has 3 branches of RQGYSP and RS which has 2 branches of RQGY. However, the improvement resulting from the bonding of the terminator to the C-end of the antiviral peptide is so great that SPC3 and RL can be reduced to two branches (SPC3 dimer and RL dimer, respectively), or even to one branch (SPC monomer and RL monomer, respectively). As these are much smaller molecules, they are much easier and cheaper to make and are preferred for that reason.

The  $\omega$ -amino-fatty acid is preferably saturated. Longer chains than 10 carbon atoms are unnecessary as the effect is obtained with less, and longer chains may be too lipidic. The preferred length is from 4 to 8 carbon atoms, and more preferably from 4 to 6 carbon atoms. The most preferred  $\omega$ -amino-fatty acids are  $\gamma$ -aminobutyric acid,  $\delta$ -aminovaleric acid and  $\epsilon$ -aminocaproic acid.

The peptidic cell membrane penetrating agent is suitably a TAT-derived peptide, penetratin® or Kpam, although other peptides may also be suitable.

### Experimental

We first synthesized SPC3 octomers, with the graft of saturated fatty acid chains of increasing length, from 4 to 8 carbons, on the core lysine residue; and SPC3 octomers with three different peptide chains on the lysine residue: a TAT-derived peptide, penetratin, and Kpam peptide, all reported to enhance membrane penetration and crossing. We tested the above molecules on C8166 cells infected with NL 4-3 HIV strain, then on PBL with the same strain.

When positive results were observed, further attempts were made to test whether the graft of membrane affinity chains on the water soluble peptides could allow for a reduction in their size without losing efficacy (SPC3, RL and their derivatives are polymers, often octomers, of small peptides: the monomers have been shown to be inactive), with a view of cost-containment. To this end we synthesized monomers and dimers of the sequences of SPC3, RL and RS, with the addition of the preferred grafted sequence, and tested them on C8166 and PBL.

## Test Methods

### *Cells and viruses.*

HIV-1 NL 4-3 isolate (Adachi et al.,1986 ; Barré-Sinoussi et al.,1983) and highly cytopathic Zairian HIV-1 NDK isolate (Ellrodt et al.,1984) was propagated in permissive CEM cells (Nara et al.,1987). Uninfected CEM and C8166 ( Salahuddin et al.,1983) were maintained in RPMI 1640 (R10) with ultraglutamine ( cambrex, Vervier, Belgium), penicillin (100 U/ml), streptomycin (100µg/ml), and 10% heat- inactivated fetal calf serum ( Cambrex).

Peripheral blood lymphocytes from an HIV-1 negative donor were grown as described earlier, maintained in RPMI 1640 with ultraglutamine, supplemented with IL2 (20 µg/ml), penicillin (100 U/ml), streptomycin (100µg/ml), and 10% heat- inactivated fetal calfserum. Cells were stimulated three days in the medium supplemented with phytohemagglutinin (20 U/ml PHA P, DIFCO, Detroit MI).

### *HIV-1 infection of C8166 cells*

Samples of  $3 \times 10^5$  /100 µL C8166 cells were preincubated in 96-well microtiter plates in culture medium containing various concentrations of peptide. After a 1 h treatment at 37°C, 100 µl of diluted viral solution of HIV-1 was added. The cells were exposed to the virus for 1 h at 37°C at a multiplicity of infection of 1000 TCID<sub>50</sub> per ml. The cells were washed three times and cultured at  $3 \times 10^5$  /ml of R10 with the treatment in 24-well plates incubated at 37°C. C8166 culture medium was replaced at Day-4 post-infection. The treatment was permanent before virus adsorption, during virus adsorption and after infection. Assays on C8166 cells have been performed at least twice and in duplicate. Toxicity was evaluated by daily cell count and trypan-blue exclusion assay. Infection of C8166 T-cells with HIV-1 was assessed by virus-induced cytopathic effects (syncytia formation) and by quantification of cell free p24 viral protein in the culture supernatants. Measurements of HIV-1 p24<sup>agg</sup> concentration in the culture supernatants were achieved by ELISA, ( ALLIANCE (R) HIV-1 p24 kit. Perkin Elmer, life sciences, USA).

### *Infection of human peripheral blood lymphocytes(PBL)*



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Samples of  $10^6$  /100  $\mu$ L PBL cells were preincubated in 96-well microtiter plates in culture medium containing various concentrations of peptides. After a 1 h treatment at 37°C, 100  $\mu$ l of diluted viral solution of HIV-1 was added. The cells were exposed to the virus for 1 h at 37°C at a multiplicity of infection of 1000 TCID<sub>50</sub> per ml. The cells were washed three times and cultured at  $10^6$  /ml of medium with the treatment in 24-well plates incubated at 37° in culture medium with the peptides in 5% CO<sub>2</sub>. The treatment was permanent before virus adsorption, during virus adsorption and after infection. The PBL culture medium was replaced every 3-4 days during three weeks always in the presence of peptide. The cell viability was assessed by cell counts and trypan-blue exclusion assay. The viral production in the culture supernatant was quantified by p24 ELISA test, as described earlier. All the experiments have been done in blind-tests. Tests have been done in duplicate.

## Results

Table 1

### Experiment on C8166 cells with HIV NL-4-3

S1	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
5 $\mu$ M	-TOX	NEG	-TOX	-TOX	-TOX	72
	-TOX	NEG	-TOX	-TOX	-TOX	73
2 $\mu$ M	-	19	-	-	-	NEG
	-	46	-	-	-	223
1 $\mu$ M	-	14	(+)	+	+	188
	-	10	-	-	-	169
0.5 $\mu$ M	-	40	-	+	+	234
	-	45	(-)	+	+	75
S2	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
5 $\mu$ M	-TOX	NEG	-TOX	TOX	TOX	ND
	-TOX	NEG	-TOX	TOX	TOX	
2 $\mu$ M	-	NEG	-	-	-	NEG
	-	NEG	-	-	-	24
1 $\mu$ M	-	NEG	-	-	-	181
	-	NEG	-	-	-	245
0.5 $\mu$ M	-	NEG	-	-	+	150
	-	NEG	-	-	+	73
S3	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
5 $\mu$ M	-	NEG	-	-	-	28
	-	NEG	-	-	-	180
2 $\mu$ M	-	NEG	-	-	-	18
	-	NEG	-	-	-	7
1 $\mu$ M	-	NEG	-	-	-	9
	-	15	-	-	+	234
0.5 $\mu$ M	-	NEG	-	-	+	97
	-	7	-	-	(+)	NEG

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Table 1 (continued)

S4	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
5 $\mu$ M	-	NEG	+	+	++	21020
	-	12	-	+	++	15674
2 $\mu$ M	-	17	-	+	++	17872
	-	14	-	+	++	24806
1 $\mu$ M	-	96	+	+	++	19801
	-	244	+	+	++	21640
0.5 $\mu$ M	-	43	+	+	++	19801
	-	28	+	+	++	25000
S5	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
10 $\mu$ M	-	NEG	-	-	-	81
	-	NEG	-	-	-	134
5 $\mu$ M	-	NEG	-	-	-	66
	-	NEG	-	-	+	71
2 $\mu$ M	-	NEG	-	-	-	206
	-	NEG	-	-	-	76
1 $\mu$ M	-	NEG	-	-	-	NEG
	-	NEG	-	-	+	152
0.5 $\mu$ M	-	NEG	-	-	-	NEG
	-	NEG	-	-	+	233
S6	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
10 $\mu$ M	-	NEG	-	-	-	NEG
	-	NEG	-	-	-	NEG
5 $\mu$ M	-	NEG	-	-	-	NEG
	-	NEG	-	-	-	NEG
2 $\mu$ M	-	NEG	-	-	-	67
	-	8	-	-	-	164
1 $\mu$ M	-	12	-	(+)	+	218
	-	25	-	-	+	186
0.5 $\mu$ M	-	75	-	+	+	1417
	-	14	-	+	++	20139
A2T	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
1 $\mu$ M	-	NEG	-	-	-	NEG
	-	NEG	-	-	-	NEG
SPC 3	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
5 $\mu$ M	-	NEG	-	-	-	NEG
	-	NEG	-	-	-	NEG
2 $\mu$ M	-	17	(+)	+	++	7589
	-	6	(+)	+	++	14787
TCell	Day 4	P 24 (pg/ml)	Day 5	Day 6	Day 7	P 24 (pg/ml)
	-	NEG	-	-	-	NEG
	-	NEG	-	-	-	NEG
NL4-3 1/1000	(+)	244	+	++	++	25000
	(+)	244	+	++	++	25000

KeyS1: SPC3-( $\eta$ -aminocaprylic acid)

S2: SPC3-Penetratin

S3: SPC3-Tat

S4 SPC3-( $\alpha$ -aminocaproic acid)S5: SPC3-( $\delta$ -aminovaleric acid)S6: SPC3-( $\gamma$ -aminobutyric acid)

++, +, (+), (+)- and - represent decreasing numbers of syncytia formed

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Table 2

## Experiment on C8166 cells with HIV NL-4-3

S2	Day 4	P-24 (pg/ml)	Day 5	Day 6	Day 7	P-24 (pg/ml)
1 $\mu$ M	-	NEG	-	-	-	67
	-	NEG	-	-	-	19
0.5 $\mu$ M	-	2	-	-	+	91
	-	2	-	-	(+)	113
0.1 $\mu$ M	(+)	75	+	+	++	596
	(+)	7	+	+	++	113
0.05 $\mu$ M	(+)	143	+	+	++	2923
	(+)	28	++	++	++	468
S3	Day 4	P-24 (pg/ml)	Day 5	Day 6	Day 7	P-24 (pg/ml)
1 $\mu$ M	-	3	-	-	-	327
	-	1	-	-	-	746
0.5 $\mu$ M	-	14	-	-	(+)	189
	-	2	-	-	+	72
0.1 $\mu$ M	(+)	61	-	+	++	787
	(+)	33	-	+	++	496
0.05 $\mu$ M	(+)	261	-	++	++	3664
	(+)	94	+	++	++	2064
S5	Day 4	P-24 (pg/ml)	Day 5	Day 6	Day 7	P-24 (pg/ml)
1 $\mu$ M	-	44	-	-	-	39
	-	18	-	-	(+)	385
0.5 $\mu$ M	-	9	-	(+)	+	39
	-	58	-	-	+	72
0.1 $\mu$ M	(+)	8	+	++	++	435
	(+)	73	-	++	++	137
0.05 $\mu$ M	(+)	66	+	++	++	3183
	(+)	33	+	++	++	2159
S6	Day 4	P-24 (pg/ml)	Day 5	Day 6	Day 7	P-24 (pg/ml)
1 $\mu$ M	-	36	-	-	-	54
	-	17	-	-	-	1668
0.5 $\mu$ M	-	14	-	(+)	+	493
	-	54	-	-	+	288
0.1 $\mu$ M	(+)	90	+	+	++	1957
	(+)	14	-	++	++	613
0.05 $\mu$ M	(+)	21	+	++	++	1271
	(+)	303	+	++	++	1022
S7	Day 4	P-24 (pg/ml)	Day 5	Day 6	Day 7	P-24 (pg/ml)
5 $\mu$ M	-	NEG	-	-	+	7
	-	NEG	-	-	++	105
1 $\mu$ M	(+)	44	+	++	++	7191
	(+)	19	+	++	++	641
AZT	Day 4	P-24 (pg/ml)	Day 5	Day 6	Day 7	P-24 (pg/ml)
1 $\mu$ M	-	NEG	-	-	-	1
	-	NEG	-	-	-	NEG
SPC-3	Day 4	P-24 (pg/ml)	Day 5	Day 6	Day 7	P-24 (pg/ml)
5 $\mu$ M	-	NEG	(+)	+	++	288
	-	NEG	(+)	+	++	342
1 $\mu$ M	(+)	35	+	++	++	3943
	(+)	11	+	++	++	1297
TCell	Day 4	P-24 (pg/ml)	Day 5	Day 6	Day 7	P-24 (pg/ml)
	-	NEG	-	-	-	NEG
	-	NEG	-	-	-	NEG
NL4-3 1/1000	(+)	303	++	++	++	25000
	(+)	184	++	++	++	25000

S1-S6 as above, S7: SPC3-Kpm

++, +, (+), (+)- and - represent decreasing numbers of syncytia formed.

All tested analogues showed an increased activity as compared to SPC3 (between 5 and 150 fold).

Similar results were obtained on PBL :

IC <sub>50</sub>	S1	S2	S3	S4	S5	S6	SPC3
	0.1 $\mu$ M	0.01 $\mu$ M	0.1 $\mu$ M	0.5 $\mu$ M	0.01 $\mu$ M	0.01 $\mu$ M	2 $\mu$ M

The best agents were S5 and S6, SPC3-( $\delta$ -aminovaleric acid) and SPC3-( $\gamma$ -aminobutyric acid) respectively, with an IC<sub>50</sub> of 0.1 to 0.01  $\mu$ M and no toxicity on cells at doses up to 10  $\mu$ M.

Table 3

*Experiment on C8166 cells with HIV NL-4-3*

Name	Formula	IC <sub>50</sub> ( $\mu$ M)
SPC3	(GPGRF) <sub>8</sub> -K <sub>4</sub> -K <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> COOH	0.5
SPC3 dimer valeric acid	(GPGRF) <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.05
SPC3 monomer	GPGRF	>10
SPC3 monomer valeric acid	GPGRF-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.02
RL	(RQYSPL) <sub>8</sub> -K <sub>4</sub> -K <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> COOH	0.01
RL dimer	(RQYSPL) <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> COOH	0.02
RL monomer	RQYSPL	0.5
RL dimer valeric acid	(RQYSPL) <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.05
RL monomer valeric acid	RQYSPL-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.05
RS	(RQYS) <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> COOH	0.1
RS monomer	RQYS	0.2
RS dimer valeric acid	(RQYS) <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.05
RS monomer valeric acid	RQYS-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.2

The above table shows that the graft of a valeric acid root on monomers of the peptides RL and SPC3 increases their activity on C8166 cells. In the case of SPC3, the activity becomes greater than that of the original polymerized peptide.

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Table 4

*Experiment on PBL with NL-4-3 strain*

Name	Formula	IC <sub>50</sub> ( $\mu$ M)	IC <sub>100</sub> ( $\mu$ M)
SPC3	(GPGRAP) <sub>8</sub> -K <sub>4</sub> -K <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> COOH	0.01	0.1
SPC3 monomer valeric acid	GPGRAP-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.02	0.1
RL	(RQGYSP) <sub>8</sub> -K <sub>4</sub> -K <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> COOH	0.005	0.1
RL dimer	(RQGYSP) <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> COOH	0.01	0.1
RL dimer valeric acid	(RQGYSP) <sub>2</sub> -K-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.005	0.05
RL monomer valeric acid	RQGYSP-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> COOH	0.01	1

The results show that monomers or dimers of the original peptides have an activity comparable to that of the octomers. SPC3 monomer valeric acid has an IC<sub>100</sub> of 0.1  $\mu$ M, as compared to 2  $\mu$ m for normal SPC3, and 0.5  $\mu$ M for SPC3 valeric acid. This is of importance as SPC3 contains 56 amino-acid residues, whereas the monomer contains only 6.

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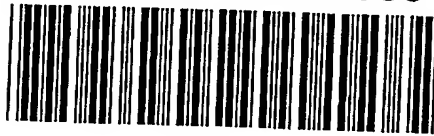
**CLAIMS**

1. A compound comprising a water soluble antiviral peptide and, bonded to the C-end thereof, a terminator which is either (a) an  $\omega$ -amino-fatty acid having from 4 to 10 carbon atoms and from 0 to 2 carbon-carbon double bonds or (b) a peptidic cell membrane penetrating agent.
2. A compound according to claim 1 in which the peptide is a multiple branch peptide construction (MBPC) with a lysine core matrix and the terminator is bonded to the root lysine residue.
3. A compound according to claim 2 in which each branch of the MBPC contains the peptide sequence GPGR.
4. A compound according to claim 3 in which each branch of the MBPC is a peptide GPGRAF.
5. A compound according to claim 2 in which each branch of the MBPC contains the peptide sequence RQGY.
6. A compound according to claim 5 in which each branch of the MBPC is a peptide RQGYSPL.
7. A compound according to claim 5 in which each branch of the MBPC is a peptide RQGYS.
8. A compound according to claim 4, claim 6 or claim 7 in which the MBPC has two branches.
9. A compound according to claim 4, claim 6 or claim 7 in which the MBPC has eight branches.
10. A compound according to claim 1 in which the peptide is GPGRAF.

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11. A compound according to claim 1 in which the peptide is RQGYSPL.
12. A compound according to claim 1 in which the peptide is RQGYS.
13. A compound according to any preceding claim in which the terminator is an  $\omega$ -amino saturated fatty acid having from 4 to 8 carbon atoms.
14. A compound according to any preceding claim in which the terminator is an  $\omega$ -amino saturated fatty acid having from 4 to 6 carbon atoms.
15. A compound according to any preceding claim in which the terminator is  $\gamma$ -aminobutyric acid,  $\delta$ -aminovaleric acid or  $\epsilon$ -aminocaproic acid.
16. A compound according to any of claims 1 to 12 in which the terminator is a TAT-derived peptide, penetratin® or Kpam.

**PCT/EP2004/005563**





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